

PAPER CHROMATOGRAPHY OF MONOMER SUGARS USING QUATERNARY SALTS FOR IDENTIFICATION

W. A. ROSENTHAL, S. SPANER AND K. D. BROWN*

*Geriatrics Research Project, Veterans Administration Hospital,
Downey, Ill. (U.S.A.)*

(Received June 6th, 1963)

INTRODUCTION

A number of procedures utilizing aniline oxalate¹, cysteine-carbazole², 2,4-dinitrophenylhydrazine³, resorcinol⁴, naphthoresorcinol⁵, orcinol⁶, vanillin⁷, *p*-anisidine¹, urea hydrochloride¹, *o*-phenylenediamine⁸, diphenylamine-aniline⁹, Benedict's solution¹⁰, ammoniacal silver nitrate¹¹ have been used for the detection of sugars on paper chromatograms. The procedures yield color differences and are sensitive to ketoses and aldoses. However, uronic acids, 5-hexulosonic acids and sugar amines fail to be selectively differentiated by the above methods.

CHARGAFF *et al.*¹² used a *m*-phenylenediamine solution and found that complexes with unsubstituted sugars possessed a marked fluorescence in the ultra-violet. ROREM¹³ has been successful in detecting and differentiating ketoses and aldoses with the aid of ultra-violet fluorescence of sulfosalicylic acid. ROREM¹⁴ was able to detect phosphorylated sugars as well as amino acids by a fluorescence technique using quinine sulfate. BERA *et al.*¹⁵ have also reported the use of cetyl-trimethyl-ammonium bromide (Cetavlon) as a spray reagent for the identification of sugars.

A major phase of our work is related to the identification of degradation products from acid mucopolysaccharides such as heparin, heparinoids, chondroitin sulfates and hyaluronic acid. A need developed during the course of our investigation for a chromagen which would permit identification of different monomer sugar classes, and subsequently different polymer fragments. This paper reports the use of organic reagents containing nitrogen which are substituted with alkyl or aryl groups capable of producing quaternary ammonium ions as possible identifying reagents. In this study, the ultra-violet fluorescence or, quenching, of the quaternary ammonium ion complex was used to detect and differentiate monomer sugars.

EXPERIMENTAL

Sample materials

The sugars or sugar derivatives used in this investigation were either obtained commercially or prepared in this laboratory. All of the samples were dissolved in water and were made up to a concentration of 1% (w/v). Multiples of one microliter aliquots were applied to the paper with a micro pipette in order to assess the sensitivity of the chromagen under investigation.

* Deceased.

Developing solvents

The following solvent systems were used:

- (A) Pyridine-ethyl acetate-acetic acid-water (5:5:1:3)¹⁶
- (B) Butanol-ethanol-water (4:1.1:1.9)
- (C) *tert.*-Amyl alcohol-propanol-ethanol-water (4:1.3:0.5:2)
- (D) Butanol-acetic acid-water (5:1.5:3.5)¹⁷
- (E) Butanol-acetic acid-water (5:1.4:2.9)¹⁷
- (F) Butanol-acetic acid-water (5:1.2:2.5)¹⁷
- (G) *tert.*-Amyl alcohol-formic acid-water (4:1:1.5)
- (H) Butanol-acetic acid-water (4.4:1.6:4.0)¹⁷

The above solvent systems are miscible at room temperature. In all cases descending chromatography was used.

Paper

Whatman No. 1 paper was found to be suitable for the procedure. Prior washing of the paper with the solvent in question was found to be beneficial, but not essential in qualitative work. Papers washed with water had little advantage over unwashed papers.

Detection

The following quaternary ammonium salts were used: (1) quinacrine hydrochloride, (2) Pyronin Y, (3) proflavine hydrochloride, (4) sparine hydrochloride, (5) methyl-dodecylbenzyl-trimethyl-ammonium chloride (Hyamine 1622), (6) cetyl-trimethyl-ammonium bromide (Cetavlon), (7) cetylpyridinium chloride, (8) cetyl-dimethylbenzyl-ammonium chloride. Each of the reagents was prepared in stock solutions containing 500 mg in 200 ml of 80% ethanol. The stock solution was diluted 1:10 with 80% ethanol for spray application.

A long wave lamp with ultra-violet radiation in the 3600Å range was used for the identification of the quaternary complexes of the sugar samples.

These lights are constructed of a special high-transmitting self-filtering glass and hence require no secondary filter. At this wavelength, the sprayed chromatographic paper shows a faint fluorescence, and the substances under analysis appear as areas of quenching with slight coloration.

Procedure

After the chromatogram has been developed, the respective solvent is removed by first air drying at room temperature, followed by drying at 100° for 3-5 min. The paper is sprayed with the desired quaternary reagent. The sprayed paper is allowed to dry at room temperature under forced air conditions. The sprayed paper is then heated for 3-5 min at 100°.

Under ultra-violet light one can locate and identify the areas of quenching.

RESULTS AND DISCUSSION

The initial phase of this investigation (*cf.* Table I) relates the sensitivity of the respective quaternary compounds to the monomer sugars. From previous work¹⁷ the

TABLE I
QUATERNARY EVALUATION
Solvent system (H): butanol-acetic acid-water (4.4:1.6:4.0)

Quaternary salt		1	2	3	4	5	6	7	8
sensitivity class		2	3	1	3	3	3	3	2
Glucose	R_F	0.47		0.46			0.50		0.45
	Color	green	none	green	none	none	white	none	white
Mannose	R_F	0.49		0.49			0.53		
	Color	lt. green	none	green	none	none	white	none	none
2,5-Anhydro-mannose	R_F	0.80		0.85			0.80		
	Color	violet	none	blue	none	none	white	none	none
Glucurone	R_F	0.49		0.53	0.53	0.55	0.55	0.54	0.54
	Color	yellow	none	orange	blue	white	white	white	white
Glucuronic acid	R_F	0.43		0.41	0.45	0.48	0.45	0.44	0.43
	Color	lt. green	none	orange	blue	white	white	white	white
Galacturonic acid	R_F	0.41		0.36	0.38	0.42		0.40	0.40
	Color	lt. green	none	orange	violet	white	none	white	white
Brucine salt of D-lyxo-5-hexuloso-nic acid	R_F	0.78	0.83	0.82	0.83	0.76	0.79	0.79	0.76
	Color	violet	violet	blue	violet	violet	violet	white	violet
Brucine salt of D-xylo-5-hexuloso-nic acid	R_F	0.79	0.82	0.82	0.83	0.77	0.79	0.80	0.77
	Color	violet	violet	blue	violet	violet	violet	white	violet
Galactosamine	R_F	0.32	0.32	0.33	0.31	0.37		0.36	0.35
	Color	lt. green	violet	violet	blue	white	none	white	white
Glucosamine	R_F	0.33	0.35	0.34	0.35	0.37		0.39	0.44
	Color	violet	violet	violet	white	white	none	white	white

butanol-acetic acid-water solvent system (H) was selected for the initial phase. The following classification of sensitivity was chosen and was used to select the most appropriate quaternary compound.

Class 1: 10–50 μg of sugar produces visually detectable level of color,

Class 2: 50–100 μg of sugar produces visually detectable level of color,

Class 3: greater than 100 μg of sugar produces visually detectable level of color.

Sugars are reported in terms of both R_F values as well as the color of the complex under ultra-violet light.

It has been demonstrated from this work that proflavin hydrochloride is the most sensitive of the reagents examined. With solvent system (H) and proflavin hydrochloride (3), the hexoses appear as green spots; the amines appear as violet spots; whereas the uronic acids and lactones appear as orange spots. The brucine salts of the 5-hexulosonic acids, however, appear as blue spots.

Table II shows the influence of solvents upon the color of the quaternary complex when viewed under ultra-violet light using proflavin hydrochloride as the identifying reagent. It should be noted that galacturonic acid, galactosamine, glucosamine and 2,5-anhydromannose respectively exhibit different colors with different solvents. It is therefore possible to resolve a mixture containing a uronic acid, a 5-hexulosonic acid and a sugar amine and a hexose. As an example, (*cf.* Table II) this could be accomplished with glucosamine, galacturonic acid, brucine salt of D-lyxo-5-hexulosonic

TABLE II
SOLVENT EVALUATION
Chromagen (3): proflavin hydrochloride (18 h development time)

<i>Solvent</i>		<i>A</i>	<i>B</i>	<i>C</i>	<i>D</i>	<i>E</i>	<i>F</i>	<i>G</i>	<i>H</i>
Glucose	R_F	0.33	0.21	0.19	0.31	0.23	0.24	0.19	0.46
	Color	green	green	green	green	green	green	green	green
Mannose	R_F	0.34	0.28	0.29	0.39	0.30	0.30	0.24	0.49
	Color	green	green	green	green	green	green	green	green
2,5-Anhydro- mannose	R_F	0.70	0.16	0.20	0.08	0.43	0.24	0.37	0.85
	Color	orange	violet	violet	violet	violet	violet	orange	violet
Glucurone	R_F	0.77	0.38	0.42	0.45	0.40	0.40	0.35	0.53
	Color	orange	orange	orange	orange	orange	orange	orange	orange
Glucuronic acid	R_F	0.32	0.24	0.25	0.30	0.22	0.21	0.18	0.41
	Color	orange	orange	orange	orange	orange	orange	orange	orange
Galacturonic acid	R_F	0.16	0.22	0.21	0.24	0.18	0.12	0.11	0.36
	Color	orange	orange	orange	orange	violet	violet	violet	orange
Brucine salt of D- xylo-5-hexuloso- nic acid	R_F	0.80	0.48	0.31	0.75	0.65	0.67	0.62	0.82
	Color	blue	blue	blue	blue	blue	blue	blue	blue
Brucine salt of D- xylo-5-hexuloso- nic acid	R_F	0.80	0.45	0.29	0.74	0.63	0.66	0.61	0.82
	Color	blue	blue	blue	blue	blue	blue	blue	blue
Galactosamine	R_F	0.26	0.06	0.18	0.22	0.08	0.11	0.09	0.33
	Color	green	green	violet	violet	violet	violet	violet	violet
Glucosamine	R_F	0.18	0.16	0.12	0.22	0.18	0.14	0.16	0.34
	Color	green	green	green	green	green	violet	green	violet

acid and glucosamine by first using solvent system (A) and chromagen (3), followed by a duplicate set of chromatograms using solvent system (E) and chromagen (3). An alternative would be to use solvent system (A) followed by solvent system (D) and again using chromagen (3) for both.

SUMMARY

A selective procedure has been developed for the identification of hexoses, sugar acids and lactones, sugar amines and 5-hexulosonic acid salts with the aid of a quaternary ammonium compound. When proflavin hydrochloride is complexed with the respective sugars, viewed under ultra-violet light, the monomers are identified by various R_F values and colors. The procedure is sensitive to 10 micrograms of sugar.

ACKNOWLEDGEMENTS

Thanks are due to Mr. J. R. HELBERT for his continuous interest and encouragement as well as for his critical reading of the manuscript; to Miss G. WILLIAMS for her technical assistance and to Mr. T. WANG for his preparation of the 5-hexulosonic acid salts.

REFERENCES

- ¹ L. HOUGH, J. K. JONES AND W. H. WADMAN, *J. Chem. Soc.*, (1950) 1702.
- ² L. UJEJSKI AND E. R. WAYGOOD, *Can. J. Chem.*, 33 (1955) 687.
- ³ R. A. GRAY, *Science*, 115 (1952) 129.
- ⁴ W. G. C. FORSYTH, *Nature*, 161 (1948) 239.
- ⁵ S. M. PARTRIDGE, *Biochem. J.*, 42 (1948) 238.
- ⁶ A. BEVENUE AND K. T. WILLIAMS, *Arch. Biochem. Biophys.*, 34 (1951) 225.
- ⁷ A. P. MACLENNAN AND H. M. RANDALL, *Anal. Chem.*, 31 (1959) 2020.
- ⁸ M. C. LANNING AND S. S. COHEN, *J. Biol. Chem.*, 189 (1951) 109.
- ⁹ R. W. BAILEY AND E. J. BOURNE, *J. Chromatog.*, 4 (1960) 206.
- ¹⁰ R. P. MURPHY, *Nature*, 185 (1960) 455.
- ¹¹ S. M. PARTRIDGE, *Biochem. J.*, 42 (1948) 238.
- ¹² E. CHARGAFF, C. LEVINE AND C. GREEN, *J. Biol. Chem.*, 175 (1948) 67.
- ¹³ E. ROREM, *Anal. Biochem.*, 1 (1960) 218.
- ¹⁴ E. ROREM, *J. Chromatog.*, 4 (1960) 162.
- ¹⁵ B. C. BERA, A. B. FOSTER AND M. J. STACEY, *J. Chem. Soc.*, (1955) 3788.
- ¹⁶ F. G. FISCHER, *Z. Physiol. Chem.*, 302 (1955) 10.
- ¹⁷ S. M. PARTRIDGE, *Biochem. J.*, 42 (1948) 238.

J. Chromatog., 13 (1964) 152-156